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(54) Recombinant plasmid containing human interferon-beta gene.

(57) Disclosed are recombinant vector plasmids containing a DNA fragment coding for human interferon- β inserted downstream from a tryptophane promoter. The plasmids are useful for transformation of microorganisms such as Escherichia coli which transformants produce human interferon- β .

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TITLE OF THE INVENTIONRECOMBINANT PLASMID CONTAINING
HUMAN INTERFERON-B GENE

5

Background of the Invention

10 The present invention relates generally to recombinant plasmids and, more specifically, to recombinant plasmids containing a DNA coding for human interferon-B (referred to as IFN-B hereinafter) and a process for producing IFN-B.

15 Expression of genes in organisms is achieved through a series of biological reactions involving synthesis of an RNA using a DNA as a template, i.e. the transcription step and synthesis of a polypeptide based on the information of mRNA, i.e. the translation step. Such recombinant DNA
20 technology has now been developed to the extent that its industrial application has become possible.

In this connection, it is important to develop a method
for the insertion of foreign genes into a plasmid vector
for the efficient synthesis of the polypeptide coded by
25 the foreign gene in a microorganism.

For the efficient expression of foreign genes in a micro-organism, particularly in *Escherichia coli*, various attempts have heretofore been made. In order to increase the rate of transcription, lactose operon promoter, tryptophan operon promoter and the like have been employed as a promoter, i.e. the initiation site for transcription with RNA polymerase. In order to increase the rate of translation, various recombinants wherein the length between the Shine-Dalgarno sequence (referred to as SD sequence hereinafter) and the initiation site of the translation is varied, generally 3 to 15 base pairs, have been prepared. See Keiichi Itakura, *Science* 198, 1056 - 1063 (1977); A.H. Seeburg et al.: *Nature* 276, 795 - 798 (1978) and J.A. Martial et al.: *Science* 205, 602 - 607 (1979). As a practical problem, however, with use of the examples mentioned above, proteins are generally produced as fused proteins consisting of two or more proteins. Although the direct production of intact proteins has been reported by David V. Goeddel et al.: *Nature*, 281, 544 - 548 (1979); Goeddel et al.: *Nature*, 287, 411 - 416 (1980) and Goeddel et al.: *Nucleic Acid Research*, 8, 4057 - 4074 (1980), the method of direct production is inconvenient because a special synthetic DNA is used as a joint to insert the foreign gene together with the initiation codon ATG for translation downstream from the promoter. Thus, a need exists for an efficient and industrially applicable method for synthesis of a polypeptide coded by a foreign gene in a microorganism. To this end, vectors have now been constructed which overcome the deficiencies of the known plasmid vectors and which are useful particularly for efficient expression of IFN-B in *Escherichia coli*.

Summary of the Invention

In accordance with the present invention, plasmid vectors are constructed which have a *Cla*I site and a *Hind*III site

downstream from the Escherichia coli tryptophane promoter (referred to as trp promoter hereinafter) and the SD sequence of tryptophane leader peptide, derived from Escherichia coli transducing phage, λ cI857trpED10. By using the two restriction sites (ClaI and HindIII) instead of a synthetic DNA as a joint, many recombinants are readily constructed wherein the length between the SD sequence and the initiation site for translation of IFN-B is varied. As a result, IFN-B is efficiently produced by culturing an Escherichia coli strain transformed with the recombinant.

Several plasmid vectors having Escherichia coli trp promoters have been reported, e.g., Norman H. Carey: Japanese Published Unexamined Patent Application No. 36500/81; R.A. Hallewell et al.: Gene 9, 27 - 47 (1980); Emtage et al.: Nature, 283, 171 - 174 (1980) and J.C. Edman et al.: Nature, 291, 503 - 506 (1981). The plasmid vectors of the present invention on the other hand have the unique characteristic of having two or more cleavage sites for restriction endonucleases selected from ClaI, HindIII, EcoRI, BamHI, PstI and the like immediately after, about the length of 0 to 20 base pairs, the SD sequence and the efficiency of translation can be increased by adjusting the length between the SD sequence and the initiation site of the foreign gene using these cleavage sites.

Brief Description of the Drawings

In the accompanying drawings, forming a part of the specification:

Fig. 1 illustrates the DNA base sequence around the tryptophane promoter and the SD sequence;

Fig. 2 is a flow chart illustrating construction of the trp portable promoter;

Fig. 3 illustrates the cleavage maps of the mild λ strain and λ trp ED phage DNA;

Fig. 4 (A) illustrates the structure of the plasmid pKYP-1 and Fig. 4 (B) illustrates that of the plasmid pKYP-5;

Fig. 5 is a flow chart illustrating construction of plasmid pKYP-5;

Fig. 6 is a flow chart illustrating construction of plasmid pKYP-10;

Fig. 7 (A) is a flow chart illustrating construction of plasmid pKYP-11 and Fig. 7 (B) illustrates the structure of plasmid pKYP-12;

Fig. 8 is a flow chart illustrating construction of the recombinant plasmid pLE-3 containing IFN- β gene; and

Fig. 9 is a flow chart illustrating construction of the recombinant plasmids pLV-1, pLVX-1 and pM λ -2 containing IFN- β gene.

Description of the Invention

In accordance with the present invention, plasmid vectors are constructed in the following manner.

The plasmid vector containing the trp promoter is prepared by cloning a DNA fragment containing the promoter and SD sequence illustrated in Fig. 1, for example, the DNA fragment from the 1st to 139th base pairs in Fig. 1 into a vector such as pBR322, pBR325, pGA22, pACYC184 and

pACYC177. The DNA sequence of the trp promoter region as is illustrated in Fig. 1, has been determined and is reported in G.N. Bennett et al.: J. Mol. Biol. 121, 113 - 137 (1978) and F. Lee et al.: J. Mol. Biol. 121, 193 - 217 (1978). Transducing phage (referred to as λ ptrp hereinafter) carrying a tryptophane operon is used as a source of DNA containing a trp promoter. Since it is difficult to cut out only the DNA fragment containing the promoter and SD sequence illustrated in Fig. 1 from the λ ptrp DNA using a restriction endonuclease, the plasmid vector having the trp promoter is constructed through the three steps shown in Fig. 2. The general method for constructing the plasmid vector is described by the following steps (a) to (d).

(a) Purification of λ ptrp DNA:

A microorganism of Escherichia coli wherein λ ptrp is lysogenized is cultured and the phages are purified from the lysate in a conventional manner to obtain λ ptrp DNAs after the induction of phages with Mitomycin C.

(b) Cloning of a trp promoter to a plasmid:

The λ ptrp DNA obtained as above is treated as follows to clone the trp operon.

(c) Preparation of a portable promoter from the trp promoter:

5 A plasmid in which a DNA fragment containing the trp promoter is cloned is isolated from the transformant mentioned above and treated with a restriction endonuclease such as HpaII, HindIII and the like to shorten the plasmid as illustrated in Fig. 6.

10 (d) Connection of two or more trp promoters:

The plasmid is treated as illustrated in Fig. 7 to construct a plasmid having two or more trp promoters.

15 Examples of the plasmid vectors containing the trp promoter obtained by the foregoing procedure are: pKYP-1 wherein a 4.6 Kb DNA fragment derived from λ trpED is inserted into pBR325; pKYP-5 wherein a 2.6 Kb DNA fragment derived from pKYP-1 is inserted into pBR322; pKYP-10
20 wherein an about 370bp DNA fragment derived from pKYP-5 is inserted into pBR322; pKYP-11 wherein an additional trp promoter is inserted into pKYP-10, yielding a total of two trp promoters; and pKYP-12 wherein an additional trp promoter is inserted into pKYP-11, yielding a total of
25 three trp promoters.

The DNA coding for IFN- β is then inserted into the plasmid vector containing the trp promoter obtained as described above to prepare a recombinant plasmid wherein the IFN- β
30 gene is inserted downstream from the trp promoter.

The gene coding for IFN- β has been cloned and the entire nucleotide sequence of the gene was determined by one of the present inventors, see T. Taniguchi et al.: Gene 10,
35 11 - 15 (1980). Furthermore, a recombinant, pTuIFNB-5, has been constructed wherein the IFN- β gene is inserted downstream from the tufB promoter of Escherichia coli which produces 800 to 2,600 units/ml IFN- β in Escherichia

coli. In the present invention, pTuIFN β -5 is preferably used as a source of IFN- β gene, but other IFN- β genes prepared by other methods may be used. An Escherichia coli transformant carrying the plasmid pTuIFN β -5 has been deposited with the American Type Culture Collection, U.S.A. under accession No. ATCC 31879.

Particularly preferred examples of the recombinant plasmids of the present invention are pLE-3 wherein the IFN- β gene is inserted into pKYP-12 and pLV-1, pLVX-1 and pMZ-2 which are derivatives of pLE-3.

Certain specific embodiments of the invention are illustrated by the following representative examples in which Examples 2 through 5 set forth specific processes for construction of the recombinant plasmids mentioned above.

Example 1

Construction of a plasmid vector having a trp promoter:

(a), Purification of tryptophane transducing phage DNA:

A λ trp phage, λ cI857trpED10 (referred to as λ trp ED hereinafter) G.F. Miazari et al.: J. Bacteriol. 133, 1457 (1978) is lysogenized in Escherichia coli JA 194 strain (F^- , λ^- , r_k^- , m_k^- , Δ trp E5, leu6), J. Carbon et al.: Recombinant Molecules p.355 (1977), Raven Press.

The resultant lysogenic strain, JA 194 (λ trp ED) is cultured at 42°C for 30 minutes to induce λ trp ED phages and prepare λ phage lysate. The λ phages are purified from the λ phage lysate by the cesium chloride equilibrium density gradient centrifugation of Yamakawa et al.:

"Chemicals of Nucleic Acids I" Tokyo Kagaku Dojin, p. 54 - 61, (1974). The λ phages are further purified by phenol

treatment and chloroform treatment according to the method of Yamakawa et al, supra, p. 62 - 65.

(b) Cloning of the trp promoter into a plasmid:

5 The cleavage map of the λ trp ED phage DNA purified as mentioned above for EcoRI and HindIII is illustrated in Fig. 3. Based on Fig. 3, cloning of the trp operon from λ trp ED phage DNA is carried out as follows. That is, 8
10 μ g of λ trp ED DNA is digested at 37°C for 2 hours with 16 units of EcoRI (product of Takara Shuzo Co., the same shall apply hereinafter) and 16 units of HindIII (product of Takara Shuzo Co., the same shall apply hereinafter) in 20 mM Tris-HCl (pH 7.5), 75 mM NaCl, 10 mM MgCl₂ and 5
15 mM dithiothreitol. Separately, 1 μ g of plasmid pBR325 DNA is digested with 2 units of EcoRI and 2 units of HindIII by the same method as mentioned above (final volume: 30 μ l). The reactions are stopped by heating at 65°C for 5
20 minutes. Then, 15 μ l each of the digests are mixed and 500 μ M (final concentration) ATP and 5 units of T4 DNA ligase (product of New England Biolabs) are added. The mixture is allowed to react at 4°C for 18 hours.
Escherichia coli C600 SF strain, Cameron et al.: Proc. Natl. Acad. Sci. 72, 3416 (1975) is transformed with the
25 plasmid mixture by the method of S. N. Cohen et al.: Proc. Natl. Acad. Sci. 69, 2110 (1972) and transformants which are resistant to ampicillin (Ap^R), resistant to tetracycline (Tc^R) and sensitive to chloramphenicol (Cm^S) are selected. Plasmid DNAs are isolated from the
30 transformants of Escherichia coli and cleaved with EcoRI, HindIII and HpaI. One of the plasmid DNAs has the structure illustrated in Fig. 4 (A) and is named pKYP-1.

35 As illustrated in Fig. 1, a TaqI cleavage site exists between 4 base pairs downstream from the SD sequence (AAGG) present downstream from the tryptophane promoter.

The plasmid pKYP-1 is digested with TaqI and EcoRI and the digest is subjected to purification by agarose gel electrophoresis to obtain a 2.6 Kb (Kilobase) DNA fragment containing the tryptophane promoter and SD sequence. The 2.6 Kb DNA fragment is cloned in a known vector, pBR322, by the method as illustrated in Fig. 5. That is, 8 µg of pBR322 is digested at 45°C for 60 minutes with 2 units of TaqI (product of Takara Shuzo Co.) in 100 µl of a reaction mixture comprising 10 mM Tris-HCl (pH 8.4), 6 mM MgCl₂, 100 mM NaCl and 6 mM 2-mercaptoethanol. After partial digestion with TaqI, the digest is subjected to low melting point agarose gel electrophoresis, Lars Wieslander: Analytical Biochemistry 98, 305 (1979), to obtain a 4.36 Kb purified DNA fragment. About 1.5 µg of the DNA fragment is completely digested at 37°C for 3 hours with 3 units of EcoRI. About 1.0 µg of a 4.33 Kb DNA fragment is recovered by the same low melting point agarose gel electrophoresis as mentioned above. Then, 12 µg of pKYP-1 DNA is partially digested with 3 units of TaqI, about 2 µg of the resultant 8.5 Kb DNA fragment is purified by low melting point agarose gel electrophoresis and the DNA is completely digested with EcoRI by the same procedure as above to obtain about 0.5 µg of a purified 2.6 Kb DNA fragment. Then, 0.4 µg of the thus obtained 4.33 Kb DNA of pBR322 and 0.25 µg of the 2.6 Kb DNA fragment of pKYP-1 are added to 20 µl of a reaction solution consisting of 20 mM Tris-HCl (pH 7.6), 10 mM MgCl₂ and 10 mM dithiothreitol. Then, 0.5 mM ATP and 4 units of T4 DNA ligase are added to the mixture and reaction is carried out at 4°C for 18 hours. Escherichia coli C600 SF8 strain is transformed with the plasmid DNA obtained as above. The plasmids in the transformant having Ap^R and Tc^R are isolated. The plasmid DNA is digested with 6 restriction endonucleases, EcoRI, HindIII, ClaI (product of Boehringer Mannheim GmbH), HpaI (product of Takara Shuzo Co.), HincII (product of Takara Shuzo Co.)

and BamHI (product of Takara Shuzo Co.) to analyze the structure of the plasmid. The plasmid is recognized to have the structure illustrated in Fig. 4 (B) and is named pKYP-5.

5 (c) Preparation of a portable promoter from the trp promoter:

10 The plasmid vector pKYP-5 mentioned above is applicable as a DNA introducing vector since it has a ClaI site and a HindIII site on the DNA of 1 to 20 base pairs downstream from the SD sequence. Since another ClaI site is present in pKYP-5 DNA besides the ClaI site immediately after the SD sequence and the fragment with the trp promoter obtained by cutting pKYP-5 DNA with EcoRI and HindIII is a
15 little too large, i.e. 2.65 Kb, pKYP-5 is improved by the process as illustrated in Fig. 6 to obtain a plasmid having a DNA fragment containing a shorter tryptophane promoter. That is, pKYP-5 DNA is digested with HpaII and HindIII and the digest is purified to obtain a DNA
20 fragment of about 340 bp. The fragment is inserted into the pBR322 digested with ClaI and HindIII as illustrated in Fig. 6 to obtain pKYP-10. The structure of pKYP-10 is determined by digestion with EcoRI, ClaI, HindIII and HpaI and agarose gel electrophoresis.

25 (d) Connection of two or more trp promoters:

30 In order to construct a plasmid vector having a stronger promoter activity, two trp promoters are inserted into the vector. That is, the DNA fragment of about 340 bp containing the trp promoter mentioned in step (c) above is inserted into the pKYP-10 digested with ClaI and HindIII to obtain pKYP-11 as illustrated in Fig. 7 (A). The same procedure is repeated to construct pKYP-12 illustrated in
35 Fig. 7 (b) wherein three trp promoters are connected at the same orientation. The structure of pKYP-12 is

confirmed by digestion with EcoRI, ClaI, HindIII and HpaI.

5 Plasmid vectors, pKYP-5, 10, 11 and 12 mentioned above are
plasmid vectors wherein the DNA fragment containing the trp
promoter is cloned in the plasmid pBR322. Other plasmids,
such as pBR325, may also be employed for the construction
of similar DNA introducing vectors. Analogous derivatives
10 of pKYP-5, 10, 11 and 12 can be constructed by deleting
the ampicillin-resistant gene or tetracycline-resistant
gene or introducing other drug-resistant genes such as
kanamycin-resistant gene, chloramphenicol-resistant gene
and the like.

Example 2

15 Insertion of the DNA coding for IFN- β into the plasmid
vector pKYP-12:

Plasmid pTuIFN β -5, isolated from ATCC 31879, by
conventional techniques, is digested with HindIII. The
20 digest is treated with DNA polymerase I (product of New
England Biolabs) and subjected to ligation to obtain
pTuIFN β H-5 illustrated in Fig. 8. The IFN- β gene
recovered from pTuIFN β H-5 is then cloned in the plasmid
pKYP-12 having a trp promoter. That is, 2 μ g of pKYP-12
25 DNA is digested with 4 units of ClaI in 30 μ l of a buffer
solution (referred to as Cla⁻ buffer hereinafter)
consisting of 10 mM Tris-HCl (pH 7.5), 6 mM MgCl₂ and 5
mM dithiothreitol at 37°C for 2 hours. NaCl is added to a
final concentration of 100 mM and the reaction is
30 continued at 37°C for 2 hours. The mixture is heated at
65°C for 5 minutes to inactivate the enzyme and is then
subjected to low melting point agarose gel electrophoresis
to obtain 1.2 μ g of a purified DNA of about 5 Kb
containing the trp promoter. Separately, 15 μ g of
35 pTuIFN β H-5 DNA is digested with ClaI and BamHI and
purified by low melting point agarose gel electrophoresis

as described above to obtain about 1 μ g of a DNA fragment (1.1 Kb) containing the IFN- β gene. The two DNA fragments obtained as above and illustrated as 5 Kb and 1.1 Kb in Fig. 8 are dissolved in a mixture of 20 mM Tris-HCl (pH 7.5), 6 mM $MgCl_2$, 5 mM dithiothreitol and 500 μ M ATP. 4 units of T4 DNA ligase is added and the mixture is allowed to react at 4°C for 18 hours. *Escherichia coli* HB101 is transformed with the mixture of the thus obtained recombinants by conventional technique to obtain colonies resistant to ampicillin. Plasmids are recovered from the culture of the colonies. One of the plasmids is pLE-3 illustrated in Fig. 8. The structure of pLE-3 is confirmed by agarose gel electrophoresis after the digestion of DNA with *Cla*I, *Eco*RI, *Hind*III and *Bam*HI. It is confirmed by the method of Maxam & Gilbert, Proc. Natl. Acad. Sci. 74, 560 (1977) that the base sequence from the SD sequence, AAGG, to the initiation codon, ATG, in the plasmid pLE-3 is "AAGGGTATCGATG".

Escherichia coli containing plasmid pLE-3 has been deposited with the American Type Culture Collection, U.S.A. as *Escherichia coli* ILE-3, ATCC 39010.

Example 3

In this example, 2 μ g of the pLE-3 DNA obtained in Example 2 is dissolved in 30 μ l of the *Cla*I⁻ buffer and 4 units of *Cla*I is added. The mixture is allowed to react at 37°C for 2 hours and is then heated at 65°C for 5 minutes to inactivate *Cla*I. Then, d-GTP and d-CTP are added to a concentration of 20 μ M each and 6 units of *Escherichia coli* DNA polymerase I (1 μ l) is added. The mixture is allowed to react at 15°C for 1 hour. Then, 5 mM dithiothreitol, 500 μ M ATP and 20 units of T4 DNA ligase are added. The mixture is allowed to react at 4°C for 18 hours. *Escherichia coli* HB101 is transformed using the reaction mixture in a conventional manner to obtain an

Ap^R colony. A plasmid is recovered from the culture of the colony, named pLV-1, is illustrated in Fig. 9 (a). The structure of pLV-1 is confirmed by agarose gel electrophoresis after the digestion of DNA with EcoRI, ClaI, HindIII and BamHI. It is confirmed by the method of Maxam & Gilbert that the base sequence from the SD sequence to the initiation codon (ATG) in pLV-1 is "AAGGGTATCGCGATG".

Escherichia coli containing plasmid pLV-1 has been deposited with the American Type Culture Collection, U.S.A. as Escherichia coli ILV-1, ATCC 39025.

Example 4

Plasmid pLVX-1 is obtained from pLE-3 mentioned in Example 2 as follows. First, 3 µg of pLE-3 DNA is dissolved in 40 µl of the Cla-buffer and 6 units of ClaI is added. The mixture is allowed to react at 37°C for 2 hours and heated at 65°C for 5 minutes to inactivate ClaI. Then, d-GTP and d-CTP are added to a concentration of 20 µM each and 6 units of Escherichia coli DNA polymerase I is added. The mixture is allowed to react at 15°C for 1 hour. Then, 5 mM dithiothreitol, 500 µM ATP, 0.1 µg of Xho-Linker, d(CCTCGAGG), (product of Collaborative Res.) and 20 units of T4 DNA ligase are added and ligation is carried out at 4°C for 18 hours. Escherichia coli HB101 is transformed using the recombinant plasmid mixture by conventional technique to obtain an Ap^R colony. A plasmid is recovered from the culture of the colony, named pLVX-1, is illustrated in Fig. 9 (b). The structure of pLVX-1 is confirmed by agarose gel electrophoresis after the digestion of DNA with EcoRI, Xho-I, HindIII and BamHI. It is confirmed by the method of Maxam & Gilbert that the base sequence from the SD sequence to the initiation codon (ATG) in pLVX-1 is "AAGGGTATCGCCTCGAGGCGATG".

Escherichia coli containing plasmid pLVX-1 has been deposited with the American Type Culture Collection, U.S.A. as Escherichia coli ILVX-1, ATCC 39024.

5 Example 5

Plasmid pMZ-2 is derived from pLVX-1 obtained in Example 4 by adjusting the length from the SD sequence to the initiation codon as illustrated in Fig. 9 (c). That is, 5 µg of pLVX-1 is dissolved in 50 µl of the Cla-buffer and 10 150 mM NaCl and 10 units of XhoI are added. The mixture is allowed to react at 37°C for 2 hours and is then heated at 65°C for 5 minutes to inactivate XhoI. Then, 8 units of DNA polymerase I is added and the mixture is allowed to react at 37°C for 30 minutes to remove the cohesive end 15 formed by the digestion with XhoI. After inactivating DNA polymerase by heating at 70°C for 15 minutes, 5 mM dithiothreitol, 500 µM ATP and 30 units of T4 DNA ligase are added and the ligation reaction is carried out. Escherichia coli HB101 is transformed with the DNA 20 solution obtained above to obtain an Ap^R colony. A plasmid is recovered from the culture of the colony, named pMZ-2, is illustrated in Fig. 9 (c). The structure of pMZ-2 is confirmed by agarose gel electrophoresis after the digestion of DNA with EcoRI, HindIII, XhoI and BamHI.

25

Escherichia coli containing plasmid pMZ-2 has been deposited with the American Type Culture Collection, U.S.A. as Escherichia coli IMZ-2, ATCC 39023.

30 Example 6

Productivity of interferon by Escherichia coli HB101 strains having the recombinant plasmids pLE-3, pLV-1, pLVX-1 and pMZ-2 obtained in Examples 2 to 5, that is 35 ILE-3, ILV-1, ILVX-1 and IMZ-2, respectively is examined as follows. ILE-3, ILV-1, ILVX-1 and IMZ-2 are cultured at 37°C for 18 hours in an LG medium (pH adjusted to 7.0

with NaOH) consisting of 10 g/l trypton, 5 g/l yeast extract 5 g/l NaCl and 2 g/l glucose. Then, 0.2 ml of the culture is inoculated in 10 ml of an MCG medium (pH 7.2) consisting of 0.6% Na_2HPO_4 , 0.3% KH_2PO_4 , 0.5% NaCl, 0.1% NH_4Cl , 0.5% glucose, 0.5% casamino acid, 1 mM MgSO_4 and 4 $\mu\text{g/ml}$ vitamin B_1 . Culturing is carried out at 30°C for 4 to 8 hours. Then, 10 $\mu\text{g/ml}$ indolacrylic acid (referred to as IAA hereinafter) which is an inducer of the tryptophane gene is added and culturing is continued for an additional 5 to 12 hours. Cells are harvested by centrifugation at 8000 rpm for 10 minutes and washed with 30 mM NaCl and 30 mM Tris-HCl (pH 7.5) buffer solution. The washed cells are suspended in 1 ml of the buffer solution mentioned above and 200 μg of lysozyme and 5 μl of 0.25 M EDTA (ethylene diamine tetraacetic acid) are added. The mixture is allowed to stand at 0°C for 30 minutes. Freezing and melting are repeated three times to disrupt the cells. The disrupted cells are subjected to centrifugation at 15,000 rpm for 30 minutes to obtain a supernatant fluid. The amount of the interferon in supernatant is determined by the method of J.A. Armstrong et al.: Appl. Microbiol. 21, 723 - 725 (1971), wherein Versicular Stomatitis Virus is used as a virus and Wish cell derived from human amnion cells is used as an animal cell.

The result is shown in Table 1.

Table 1

30	Strain	Plasmid contained	β -IFN (units/l)
	ILE - 3	pLE - 3	2×10^8 - 6×10^8
	ILV - 1	pLV - 1	7×10^8 - 5×10^9
35	ILVX - 1	pLVX - 1	5×10^6 - 2×10^7
	IMZ - 2	pMZ - 2	1×10^9 - 8×10^9

WHAT IS CLAIMED IS:

1. A recombinant plasmid wherein a DNA fragment coding for human interferon- β is inserted downstream from at least one tryptophane promoter.

2. The recombinant plasmid according to claim 1, wherein two or more cleavage sites for restriction endonucleases are present within the length of 0 to 20 base pairs downstream from the SD sequence.

3. The recombinant plasmid according to claim 2, wherein the restriction endonucleases are selected from ClaI, HindIII, EcoRI, BamHI and PstI.

4. A recombinant plasmid according to claim 1 which is selected from the group consisting of plasmids pLE-3, pLV-1, pLVX-1 and pM2-2.

5. A plasmid derivative prepared by treating a plasmid according to claim 1 with a restriction endonuclease or a DNA modification enzyme selected from exonuclease III, S1 nuclease and Bal 31.

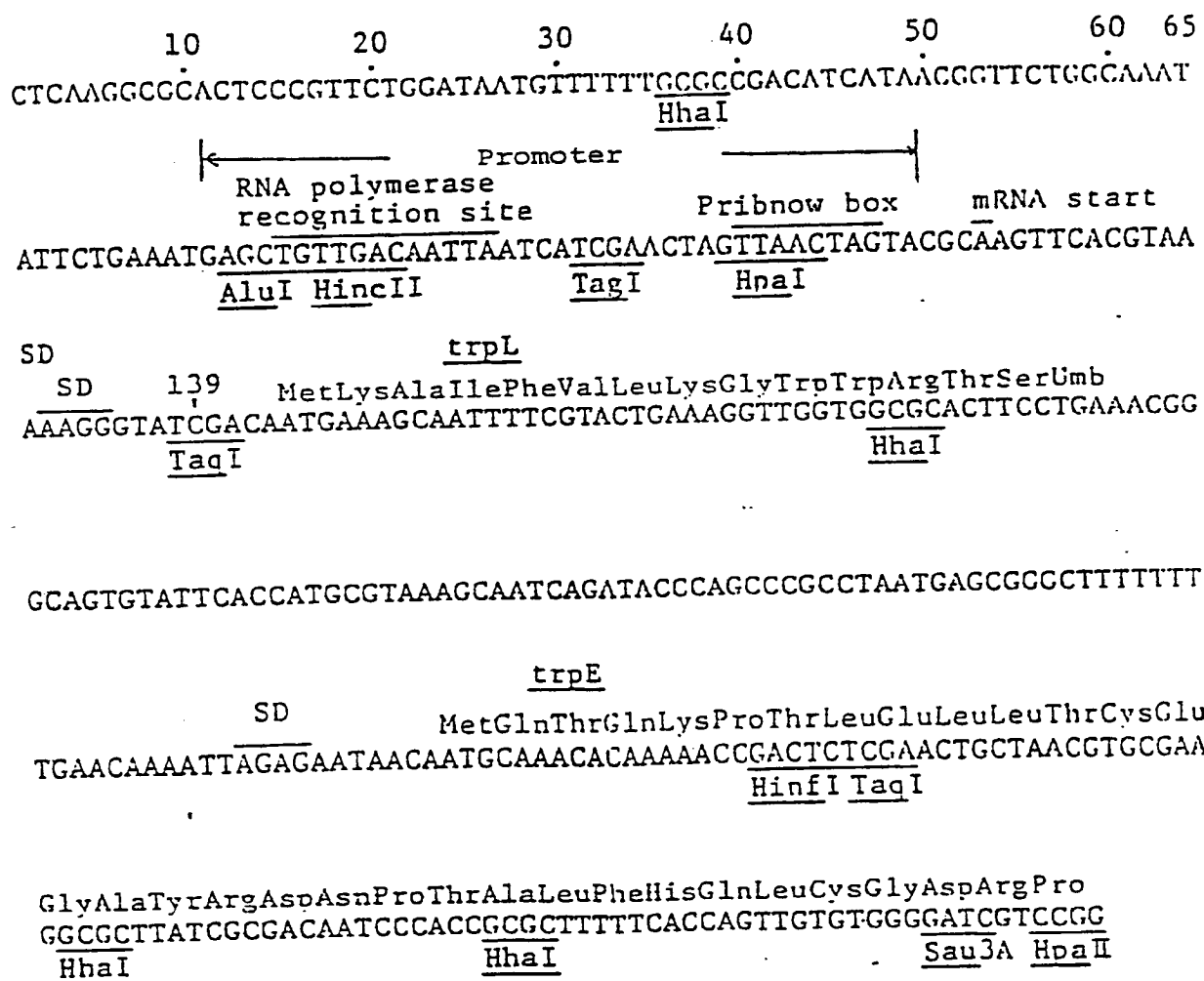
6. A process for producing human interferon- β , which comprises culturing a microorganism transformed with a recombinant plasmid as defined in claim 1, 2, 3, 4 or 5 in a medium, accumulating human interferon- β in the medium and recovering human interferon- β therefrom.

7. Microorganism transformed with a recombinant plasmid as defined in anyone of claims 1 to 5.

8. *Escherichia coli* characterized in that it contains plasmid ple-3, plv-1, plvx-1, or pmz-2.

9. Microorganism for the production of interferon characterized in that it is *Escherichia coli* ile - 3 (atcc 39010) ilv-1 (atcc 39025), ilvx-1 (atcc 39024) imz-2 (atcc 39023).

Fig. 1

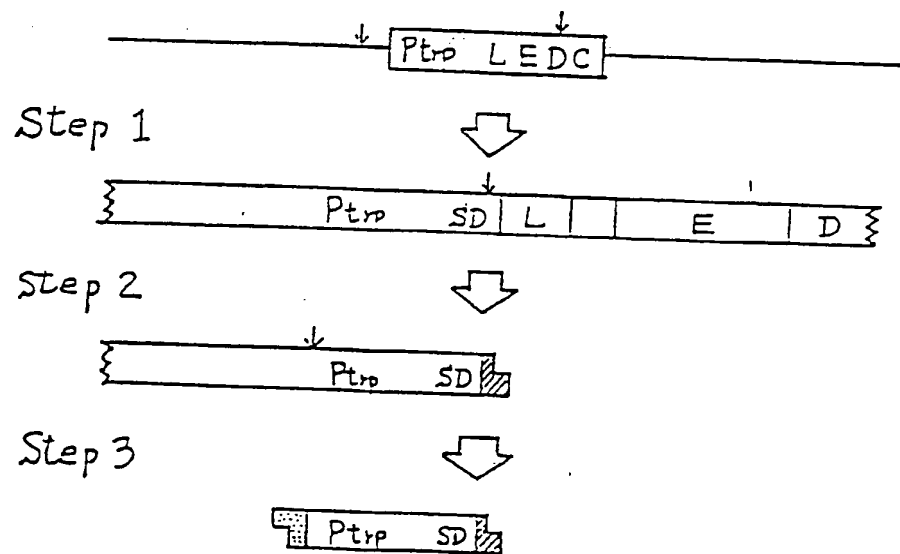


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Case 313 -

Fig. 2



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Case 313 -

Fig. 3

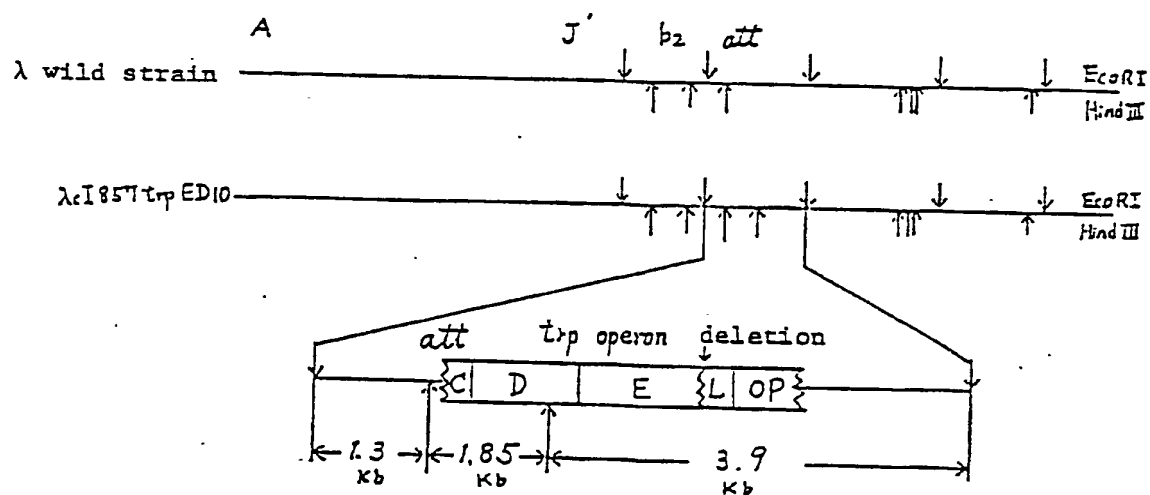
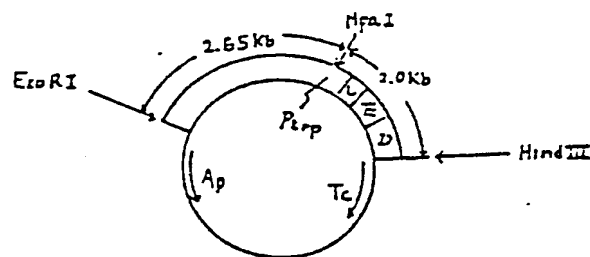


Fig. 4

(A)

pKYP-1 (9.2 Kb)

(B)

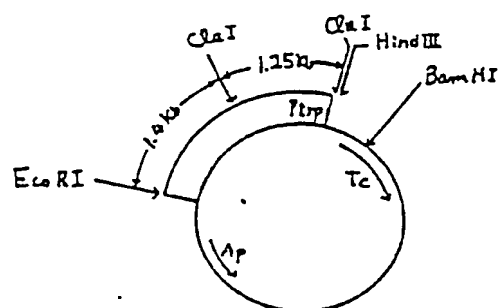
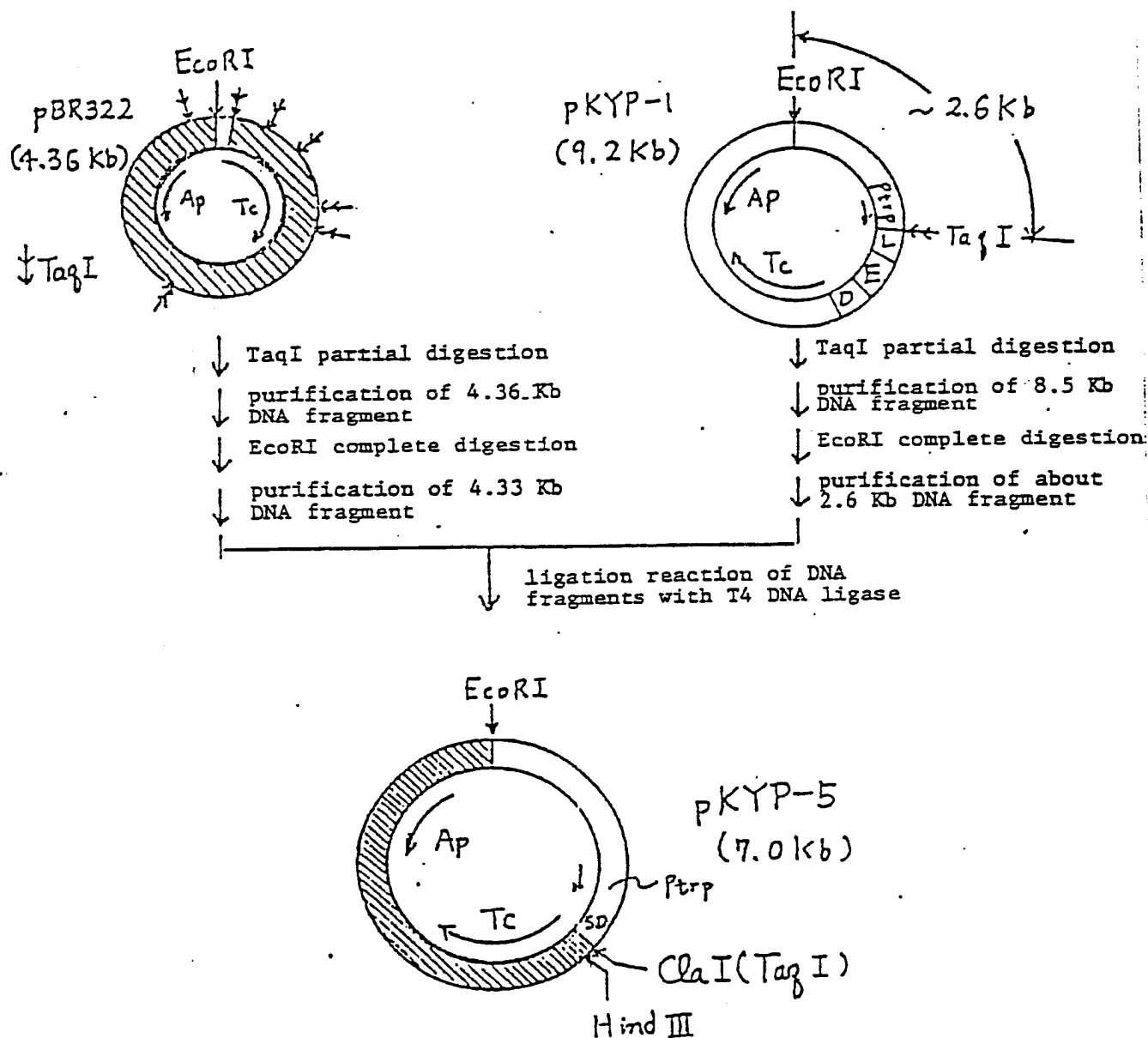
pKYP-5 (5.4 kb)

Fig. 5

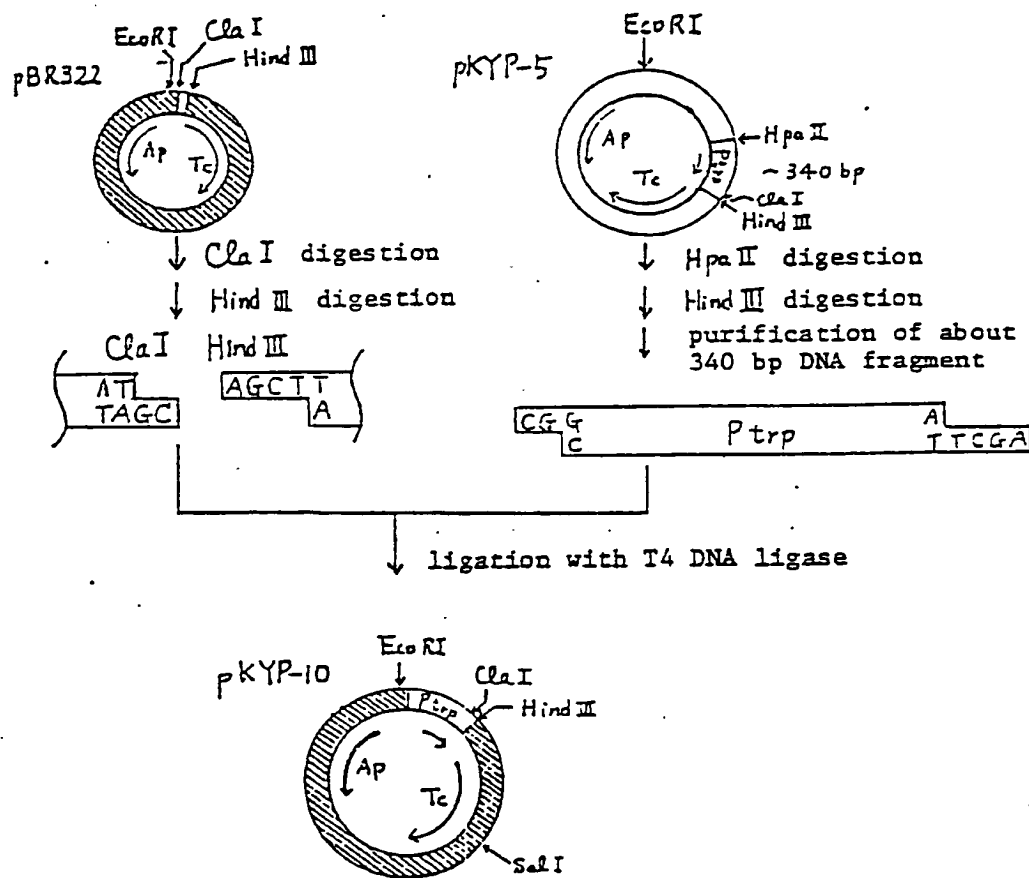


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Fig. 6

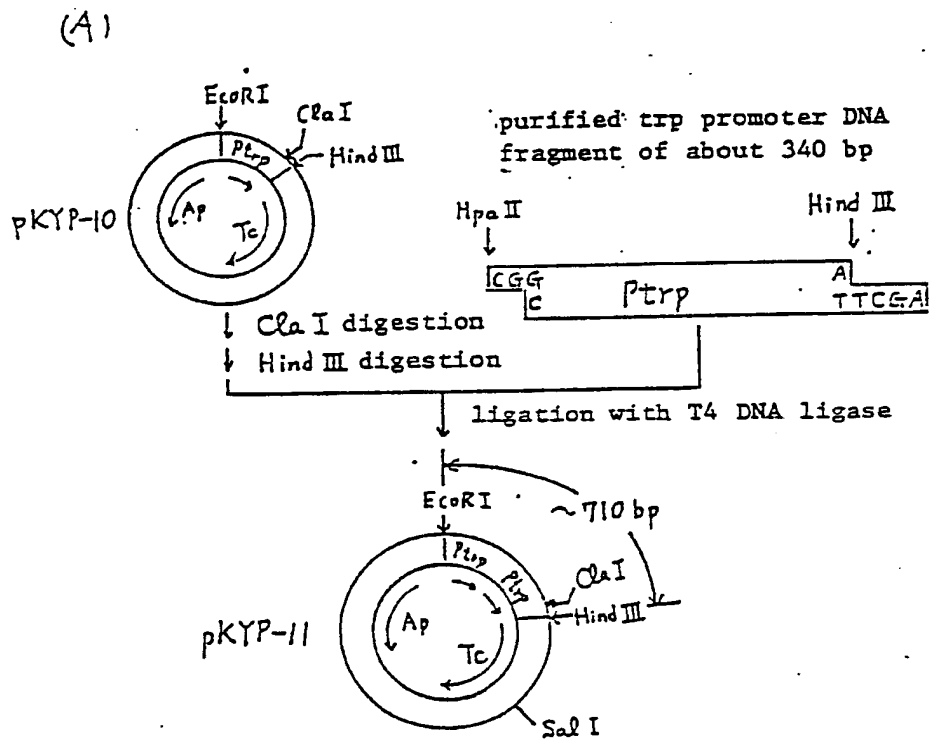


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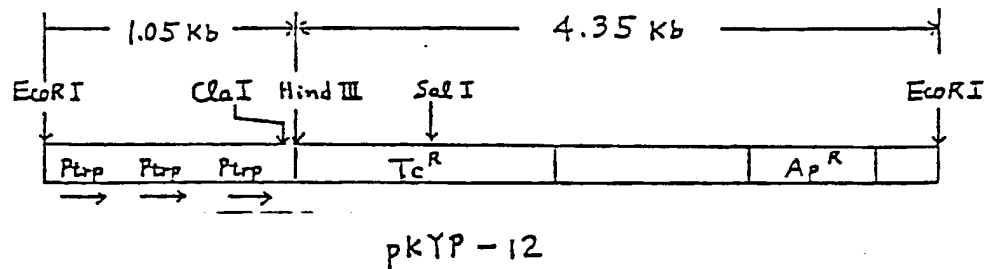
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Fig. 7



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Fig. 8

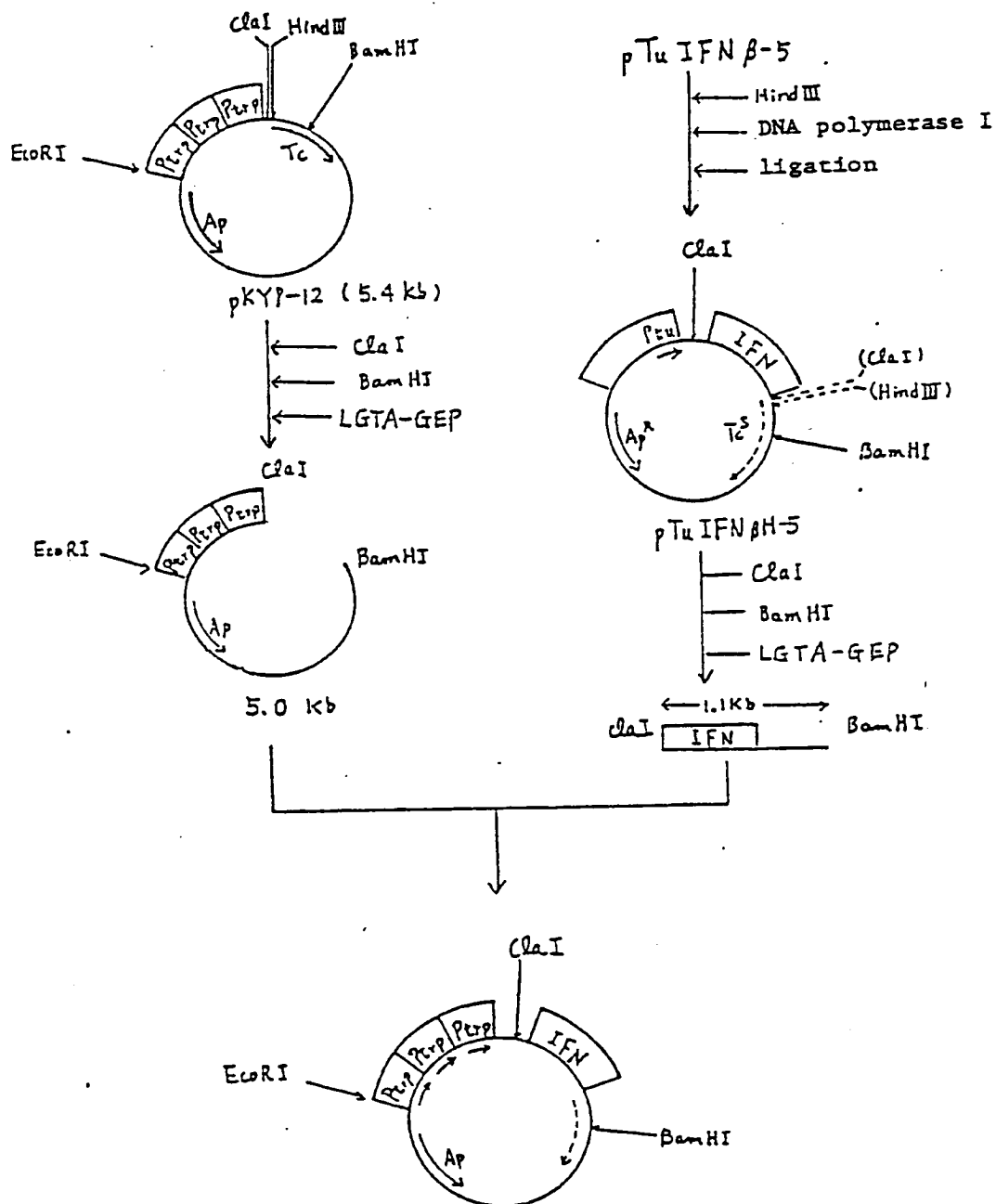
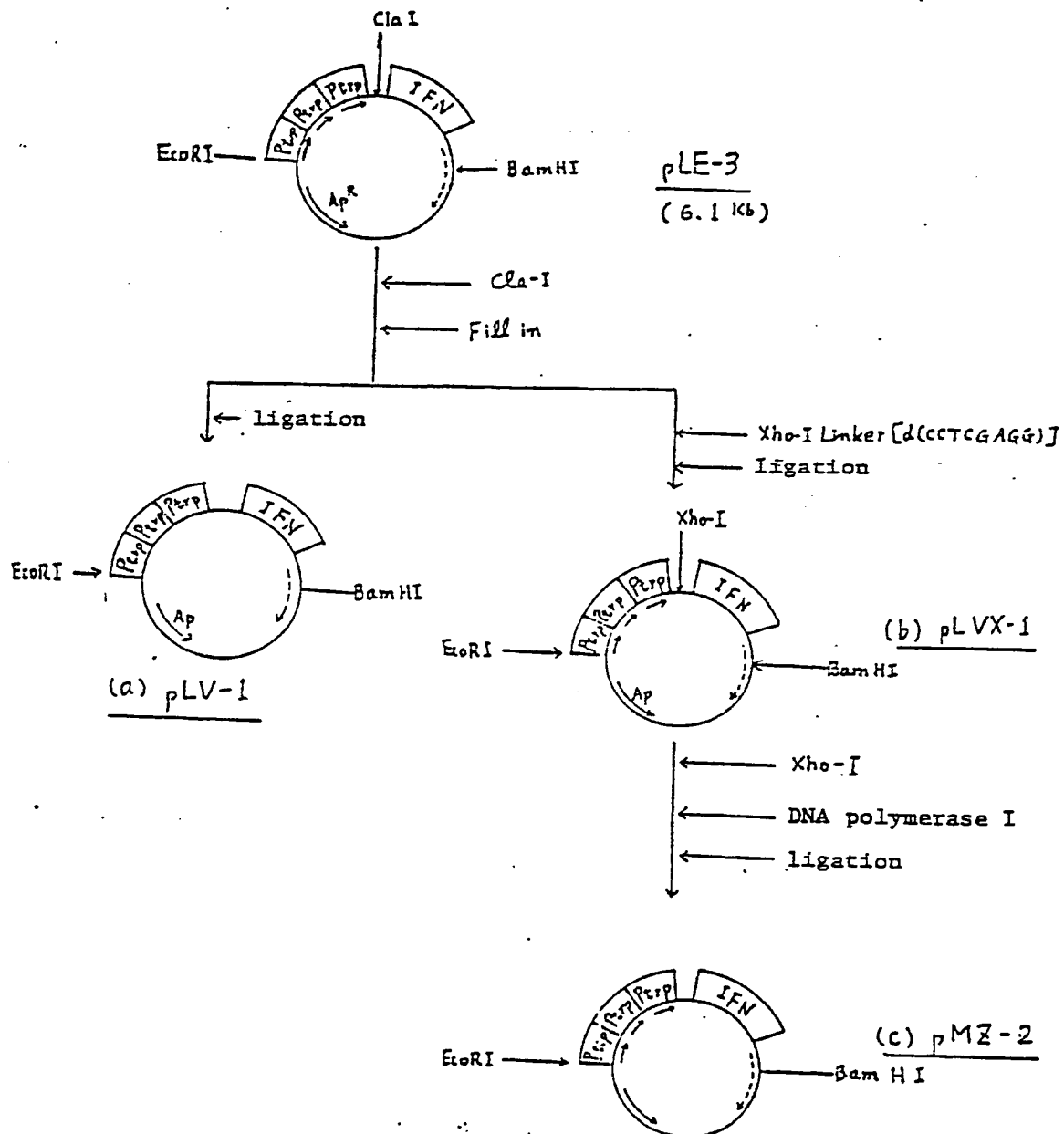


Fig. 9





European Patent Office

0083069

Application number:

82771895.7

DECLARATION PURSUANT TO RULE 28, PARAGRAPH 4,
OF THE EUROPEAN PATENT CONVENTION

The applicant has informed the European Patent Office that, until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, the availability of the micro-organism(s) identified below, referred to in paragraph 3 of Rule 28 of the European Patent Convention, shall be effected only by the issue of a sample to an expert.

IDENTIFICATION OF THE MICRO-ORGANISMS

Accession numbers of the deposits:

atcc	39010	(20-11-81)
"	39023	(14-12-81)
"	39024	(14-12-81)
"	39025	(14-12-81)

1. The first part of the document discusses the importance of maintaining accurate records of all transactions and activities. It emphasizes that proper record-keeping is essential for transparency and accountability, particularly in financial matters. The text outlines various methods for organizing and storing data, including digital databases and physical filing systems.

2. The second section focuses on the role of technology in modern record management. It highlights how software solutions can streamline processes, reduce errors, and improve accessibility. Examples of specific tools and platforms are provided, along with a discussion on the security measures necessary to protect sensitive information from unauthorized access or data breaches.

3. The third part of the document addresses the challenges associated with long-term data retention and archiving. It explores the legal requirements for preserving records and the importance of regular backups and disaster recovery plans. The text also touches upon the ethical considerations of data storage, such as privacy concerns and the potential for misuse of stored information.

4. Finally, the document concludes by summarizing the key points and offering recommendations for best practices. It encourages organizations to adopt a proactive approach to record management, ensuring that all data is properly documented, stored, and protected throughout its lifecycle. The overall goal is to provide a comprehensive guide that helps readers understand the complexities of record-keeping and implement effective strategies for their own operations.